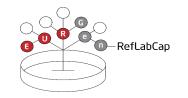
# 







# Results from the second External Quality Assessment (EQA) exercise



EURGen-RefLabCap

Faisal Ahmad Khan (fakh@food.dtu.dk)

Lauge Holm Sørensen (lahoso@food.dtu.dk)





# INTRODUCTION

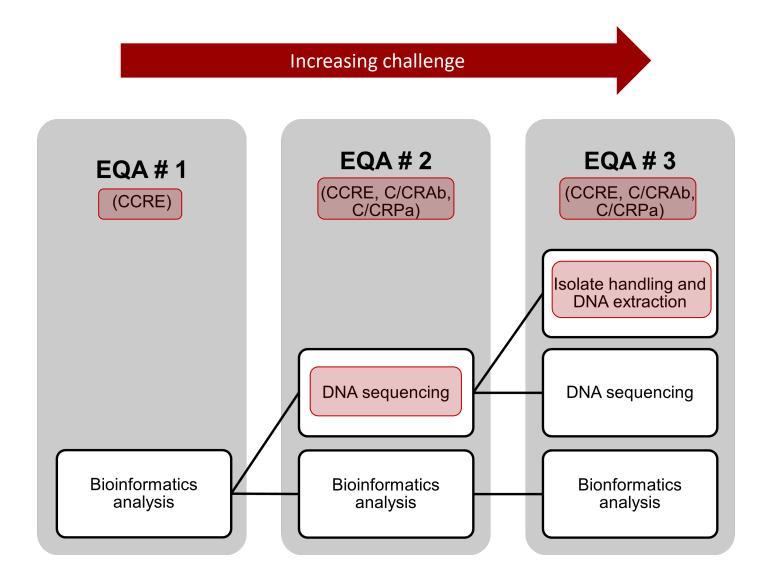
- OVERVIEW OF THE EURGen-RefLabCap EQAS
- DESIGN OF THE SECOND EQA
- PREPARATION OF EXPECTED RESULTS
- SCORING SYSTEM

\*\*\*



# OVERVIEW OF ALL EURGen-RefLabCap EQAs





- Workstream 1 pathogens (WS1)
  - CRE/CCRE
- Workstream 2 pathogens (WS2)
  - C/CRPa
  - C/CRAb

Results from second EQA exercise



# **DESIGN OF SECOND EQA**



#### **Strains:**

21-11-2023

- EURGen-2023-01 (Acinetobacter baumannii)
- EURGen-2023-02 (Escherichia coli)
- EURGen-2023-03 (Klebsiella pneumoniae)
- EURGen-2023-04 (Pseudomonas aeruginosa)

#### **Bioinformatics analyses included in EQA 2023**:

- Prediction of MLST
- 2. Prediction of plasmid replicon types
- Detetcion of genes and chromosomal mutations mediating AMR
- 4. In silico prediction of AMR profiles

#### **Materials:**

- Purified DNA
- (Raw and assembled reads from Illumina and nanopore sequencing technology)

#### **Additional Analyses**

Quality control of sequences generated by participants

- *I.* Short-read sequences
- II. Long-read sequences





## PREPARATION OF EXPECTED RESULTS



- Consensus results from TWO reference laboratories
  - Sequencing and bioinformatics analysis at DTU
  - Sequencing and bioinformatics analysis at SSI
- Bioinformatics tools used to prepare expected results
  - Mainly CGE tools
  - AMRFinder+

21-11-2023

- RGI (CARD database)
- PathogenWatch
- Default thresholds (≥80% ID and ≥ 60% COV)

#### **Final set of expected results**

Categorical agreement

Coverage ≥ 90% (plasmids)/≥ 60% (ARGs)

#### **Expected non mandatory results**

- No consensus between reference labs
- Detetcion in only one type of dataset (Ir or sr)
  - Detection in only one tool

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# SCORING SYSTEM IN THE WEBTOOL



Analysis	Submitted result	Score
Prediction of MLST	Correct MLST	1
Prediction of MLS1	Incorrect MLST	0
	Genetic determinant correctly identified	1
Detection of plasmid replicons, AMR genes	Reporting a genetic determinant that was part of the expected results but not mandatory to report	blank
and chromosomal mutations	Missing a genetic determinant	blank
	Reporting an unexpected genetic determinant	0
	AMR profile correctly reported for the antimicrobial	1
In-silico AMR profiles	Reporting an antimicrobial that was part of the expected results but not mandatory to report, or part of intrinsic resistance	blank
	Missing an antimicrobial	blank
	Reporting an AMR profile for an unexpected antimicrobial	0

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# **SCORING SYSTEM**



# **Maximum possible score of participants**

Material and analysis	EURGen- 2023-01	EURGen- 2023-02	EURGen- 2023-03	EURGen- 2023-04	Total
Prediction of MLST	1	1	1	1	4
Detection of plasmid replicons	0	6	1	0	7
Detection of AMR genes and chromosomal PMs	6	17	4	6	33
In silico prediction of AMR profiles	10	18	5	10	43
Total	17	41	12	17	87





# **RESULTS AND DISCUSSION**

- MATERIAL ANALYSED BY PARTICIPANTS
- OVERALL SCORES OF THE PARTICIPANTS
- PREDICTION OF MLST
- DETETCION OF PLASMID REPLIOCN GENES
- DETETCION OF AMR GENES AND MUTATIONS
- IN SILICO PREDICTION OF AMR PROFILES

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# MATERIAL ANALYSED BY PARTICIPANTS



**Total submission: 30** 

WS1: 30 laboratories submitted results

WS2: 28 laboratories submitted results

	EURGen-2023-01	EURGen-2023-02	EURGen-2023-03	EURGen-2023-04
	(A. baumannii)	( <i>E. coli</i> )	(K. pneumoniae)	( <i>P. aeruginosa</i> )
Number of Laboratories	28	30	30	28

#### **Test material used:**

DNA: 22 laboratories

FASTQ: 5 laboratories

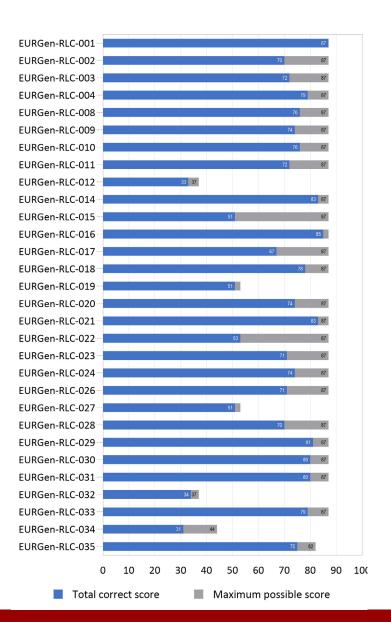
FASTA: 2 laboratories

DNA/FASTQ: 1 laboratory



# **OVERALL SCORES OF THE PARTICIPANTS**





#### **Averages of scores (%)**

Prediction of MLST: 90%

Detection of plasmid replicons: 81.9%

Prediction of genetic AMR determinants: 86%

Prediction of AMR profiles: 86.1%

Total: 85.5%





# PREDICTION OF MLST – EXPECTED RESULTS



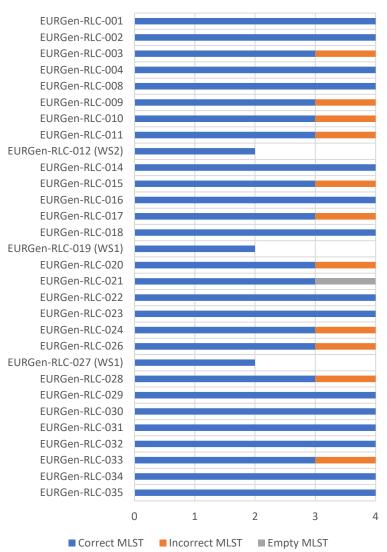
Material	MLST	Alleles assigne	Alleles assigned to each loci					
EURGen-2023-01 (A. baumannii)	136ª	cpn60	gdhB	gltA	gpi	gyrB	recA	rpoD
Lorden-2023-01 (A. baumanni)	130-	2	3ª	1	16	3	2	3
EURGen-2023-02 ( <i>E. coli</i> )	410	adk	fumC	gyrB	icd	mdh	purA	recA
EURGEII-2025-02 (E. COII)	410	6	4	12	1	20	18	7
FURCER 2022 02 (V. programonico)	4568	gapA	infB	mdh	pgi	phoE	гроВ	tonB
EURGen-2023-03 (K. pneumoniae)	4506	2	1	2	1	247	1	46
EURGen-2023-04 ( <i>P. aeruginosa</i> )	233	acsA	aroE	guaA	mutL	nuoD	ppsA	trpE
	233	16	5	30	11	4	31	41

<sup>&</sup>lt;sup>a</sup> The Oxford scheme reports two sequence types for EUGen-2023-01 due to presence of multicopy *gdhB* allele i.e., *gdhB\_*189 and *gdhB\_*3. The allele with lowest number was selected for the expected results.



# PREDICTION OF MLST - SUBMITTED RESULTS





- Average score : 90%
- All MLST were correct for EURGen-2023-02, EURGen-2023-03, and EURGen-2023-04
- 11 Incorrect MLST
  - All for EURGen-2023-01

WS1: Only submitted results for workstream 1 pathogens; WS2: Only submitted results for workstream 2 pathogens.





# PREDICTION OF MLST - DISCUSSION



#### 11 Incorrect MLST results for *A. baumannii* (EURGen-2023-01)

- Due to MLST scheme used
  - Participants used Pasteur scheme (ST 02)
  - Oxford scheme used for expected results (ST 136)
- Due to presence of multicopy gdhB allele i.e., gdhB\_3 and gdhB\_189 (Oxford scheme)
  - Resulting in two sequence types i.e., ST 136 and ST 1851
  - Also detected in expected results

- For the self-evaluation, it should be considered that these discrepancies do not represent a flaw in the bioinformatics analysis performed
- It is important to understand that the bioinformatics capacity and knowledge required for using either MLST scheme is the same

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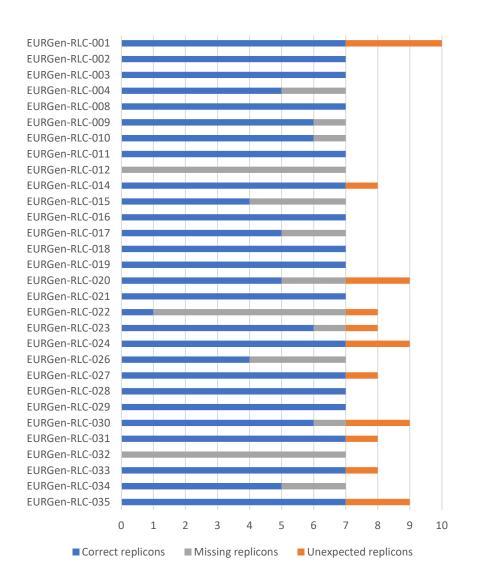
# DETECTION OF PLASMID REPLICONS – EXPECTED RESULTS

Material	Plasmid replicons	Nr.
EURGen-2023-01 (A. baumannii)	No plasmid replicon detected	0
EURGen-2023-02 ( <i>E. coli</i> )	Expected: Col(BS512), ColKP3, IncFIA, IncFIB(AP001918), IncFII(pAMA1167-NDM-5), IncX3	6
EURGen-2025-02 (E. Com)	Expected but non-mandatory: IncQ1, Col(pHAD28)	2
ELIPCon 2022 02 (V. nnoumaniae)	Expected: repB(R1701)	1
EURGen-2023-03 (K. pneumoniae)	Expected but non-mandatory: Col(pHAD28)	1
EURGen-2023-04 (P. aeruginosa)	No plasmid replicon detected	0





# DETECTION OF PLASMID REPLICONS - SUBMITTED RESULTS



#### **Total submissions: 60**

- 50% submissions were fully correct (n=30)
- 33.3% had missing replicons (n=20)
- 23.3% had unexpected replicons (n=14)

6.7% Simultaneous (n=4)

Average score: 81.9%

17 participants achieved 100% of max. possible score





# **DETECTION OF PLASMID REPLICONS - SUBMITTED RESULTS**

Strain	Missing expected replicons	Unexpected replicons
	Col(pHAD28)* (n=24)	ColpVC (n=3)
	IncQ1* (n=17)	IncFII(pRSB107) (n=2)
	ColKP3 (n=8)	IncFIA(HI1) (n=1)
EURGen-2023-02	IncFIB(AP001918)(n=7)	
	IncFII(pAMA1167-NDM-5) (n=5)	
	IncX3 (n=3)	
	Col(BS512) (n=3)	
EURGen-2023-03	Col(pHAD28)* (n=24)	Col(MG828) (n=8)
	repB(R1701) (n=8)	Col440I (n=3)

<sup>\*</sup> Expected but non-mandatory



# **DETECTION OF PLASMID REPLICONS - DISCUSSION**



- The non-mandatory replicons were missing in most results
  - Col(pHAD28) (n=43) and IncQ1 (n=17)
  - IncQ1 was only detected in long-reads sequencing data



#### <u>Discrepancy between short-and long-read data:</u>

- long-read sequencing is overall more adequate for detection of plasmids
- the assembly process might fail to properly capture sequences that were present in raw data

- The missing plasmid replicons:
  - Choice of different thresholds (?)



#### **Different approaches according to purpose:**

- Thresholds can be adjusted for different analyses
- Perhaps better to be less strict and manually evaluate results



# DETECTION OF GENES AND MUTATIONS MEDIATING AMR – EXPECTED RESULTS



Material	AMR genes and chromosomal mutations			
ELIDCon 2022 01 / A haumannii)	Expected: aph(3')-VIa, armA, bla <sub>NDM-1,</sub> bla <sub>OXA-23,</sub> gyrA S81L, parC S84L			
EURGen-2023-01 ( <i>A. baumannii</i> )	Expected but non-mandatory: ftsl A515V, parC V104I, parC D105E			
EURGen-2023-02 ( <i>E. coli</i> )	<b>Expected:</b> $aac(3)$ -IId, $aac(6')$ -Ib- $cr^b$ , $bla_{CMY-2}^c$ , $bla_{OXA-181}$ , $bla_{NDM-5}$ , $bla_{TEM-1}^d$ , $bla_{CTX-M-15}$ , $qnrS1$ , $sul1$ , $sul2$ , $dfrA12$ , $dfrA17$ , $glpT$ E448K, $gyrA$ D87N, $gyrA$ S83L, $parE$ S458A, $parC$ S80I			
	Expected but non-mandatory: bla <sub>OXA-1,</sub> ftsl N337NYRIN, pmrB Y358N			
FUDCon 2022 02 // pnoumonico)	Expected: $bla_{SHV-1}^{e}$ , $bla_{TEM-1}^{d}$ , $bla_{CTX-M-3}$ , $qnrS1$			
EURGen-2023-03 (K. pneumoniae)	Expected but non-mandatory: mgrB::IS1			
FUDCon 2022 04 /D gorusinass)	Expected: aac(6')-II, aac(3)-Id, bla <sub>VIM-2</sub> , bla <sub>OXA-4</sub> , crpP, gyrA T83I			
EURGen-2023-04 ( <i>P. aeruginosa</i> )	Expected but non-mandatory: parC S87L			

<sup>&</sup>lt;sup>a</sup> Either aph(3')-VIa or aph(3')-VI

\* \* \* \* \* \* \*

b Either aac(6')-Ib-cr5 or aac(6')-Ib-cr

<sup>&</sup>lt;sup>c</sup> Either the *bla*<sub>CMY-2</sub> or *bla*<sub>CMY-59</sub>

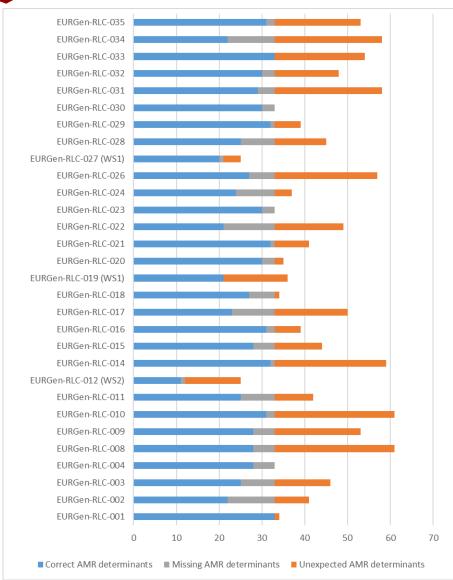
d Either bla<sub>TEM-1A</sub>, bla<sub>TEM-1B</sub>, bla<sub>TEM-1C</sub> or bla<sub>TEM-1D</sub>

e Either bla<sub>SHV-1</sub>, bla<sub>SHV-185</sub> or bla<sub>SHV-187</sub>









#### **Total submissions: 114**

- 9.6% submissions were fully correct (n=11)
- 61.4% had missing AMR determinants (n=70)
- 74.6% had unexpected determinants (n=85)

-45.6% Simultaneous (n=52)

#### Average score: 86%

- Three participants achieved 100% of their max. possible score







# DETECTION OF GENES AND MUTATIONS – SUBMITTED RESULTS

Strain	Total expected	Correct	Missing	Unexpected	
EURGen-2023-01	168	132 (78.6%)	36 (21.4%)	73 (个 43.6%)	<b>-</b>
EURGen-2023-02	493	436 (88.4%)	57 (11.6%)	76 (个15.4%)	
EURGen-2023-03	116	108 (93.1%)	8 (6.9%)	104 (个89.7%)	←
EURGen-2023-04	168	134 (79.8%)	34 (20.2%)	125 (个74.4%)	<b></b>





# DETECTION OF GENES AND MUTATIONS - DISCUSSION

Strain	Examples of problems		
	Missing gyrA S81L (n=16) or parC S84L (n=15)		
01	Unexpected $bla_{OXA-66}$ (n=15) or $bla_{ADC-25}$ (n=12)		
	Unexpected aph(3")-Ib (n=12), aph(6)-Id (n=12), tet(B) (n=12)		
	Missing glpT E448K (n=19)		
02	Unexpected aph(3")-Ib (n=14), aph(6)-Id (n=13), aadA2 (n=12), aadA5 (n=12), tet(B) (n=12)		
	Missing mgrB::IS1 (n=29)		
03	Unexpected fosA (n=19), oqxA/oqxB (n=16)		
	Unexpected acrR mutations (n=42)		
	Missing gyrA T83I (n=12)		
04	Unexpected $bla_{PAO}$ (n=21), $bla_{OXA-486}$ (n=12), $fosA$ (n=20)		
	Unexpected aadA2 (n=11), aph(3')-IIb (n=12), sul1 (n=12), dfrB5 (n=14)		

#### Missing mutations due to lacking database

- PointFinder can't detect PMs in A. baumannii
  and P. aeruginosa No database!
- glpT mutations not present in PointFinder database (ResFinder)
- mgrB::IS1, was not possible to report in webtool



#### Multiple tools and database can be used:

- AMRFinder+
- CARD
- ResFinder
- Other tools (PathogenWatch?)

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Strain	Examples of problems
	Missing gyrA S81L (n=16) or parC S84L (n=15)
01	Unexpected $bla_{OXA-66}$ (n=15) or $bla_{ADC-25}$ (n=12)
	Unexpected aph(3")-Ib (n=12), aph(6)-Id (n=12), tet(B) (n=12)
	Missing glpT E448K (n=19)
02	Unexpected aph(3")-Ib (n=14), aph(6)-Id (n=13), aadA2 (n=12), aadA5 (n=12), tet(B) (n=12)
	Missing mgrB::IS1 (n=29)
03	Unexpected fosA (n=19), oqxA/oqxB (n=16)
	Unexpected acrR mutations (n=42)
	Missing <i>gyrA</i> T83I (n=12)
04	Unexpected $bla_{PAO}$ (n=21), $bla_{OXA-486}$ (n=12), $fosA$ (n=20)
	Unexpected aadA2 (n=11), aph(3')-IIb (n=12), sul1 (n=12), dfrB5 (n=14)

#### **Reporting intrinsic resistance genes**

 Present in the strains but do no contribute to the elevated resistance in non-WT phenotype



#### **Results must be evaluated critically:**

- Too much noise can hide the important information
- Insufficient knowledge regarding genetic mechanisms of AMR might lead to incorrect reporting of resistance profiles







Strain	Examples of problems
	Missing gyrA S81L (n=16) or parC S84L (n=15)
01	Unexpected $bla_{OXA-66}$ (n=15) or $bla_{ADC-25}$ (n=12)
	Unexpected aph(3")-Ib (n=12), aph(6)-Id (n=12), tet(B) (n=12)
	Missing glpT E448K (n=19)
02	Unexpected aph(3")-Ib (n=14), aph(6)-Id (n=13), aadA2 (n=12), aadA5 (n=12), tet(B) (n=12)
	Missing mgrB::IS1 (n=29)
03	Unexpected fosA (n=19), oqxA/oqxB (n=16)
	Unexpected acrR mutations (n=42)
	Missing gyrA T83I (n=12)
04	Unexpected $bla_{PAO}$ (n=21), $bla_{OXA-486}$ (n=12), $fosA$ (n=20)
	Unexpected aadA2 (n=11), aph(3')-IIb (n=12), sul1 (n=12), dfrB5 (n=14)

#### Reporting AMR genes for antimicrobials not included in EQA

- These genes are present in the strains but they confer resistance to antimicrobials not relevant for the species
- Tetracycline (tet(B), tet(G)), Streptomycin (aadA2, aadA5, aph(6)-Id, aph(3'')-Ib) and Kanamycin (aph(3')-IIb).



#### Results must be evaluated critically:

- Results from the bioinformatics tools should not be reported without critical evaluation
- Carefully report AMR determinants







Strain	Examples of problems
	Missing gyrA S81L (n=16) or parC S84L (n=15)
01	Unexpected $bla_{OXA-66}$ (n=15) or $bla_{ADC-25}$ (n=12)
	Unexpected aph(3")-Ib (n=12), aph(6)-Id (n=12), tet(B) (n=12)
	Missing glpT E448K (n=19)
02	Unexpected aph(3")-Ib (n=14), aph(6)-Id (n=13), aadA2 (n=12), aadA5 (n=12), tet(B) (n=12)
	Missing mgrB::IS1 (n=29)
03	Unexpected fosA (n=19), oqxA/oqxB (n=16)
	Unexpected acrR mutations (n=42)
	Missing gyrA T83I (n=12)
04	Unexpected $bla_{PAO}$ (n=21), $bla_{OXA-486}$ (n=12), $fosA$ (n=20)
	Unexpected aadA2 (n=11), aph(3')-IIb (n=12), sul1 (n=12), dfrB5 (n=14)

# Reporting PMs (and genes) with unconfirmed impact on the AMR profiles

PMs in acrR and ramR



#### **Results must be evaluated critically:**

 Might highlight the insufficient knowledge of genetic AMR mechanisms

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# IN SILICO PREDICTION OF AMR PROFILES — EXPECTED REULTS

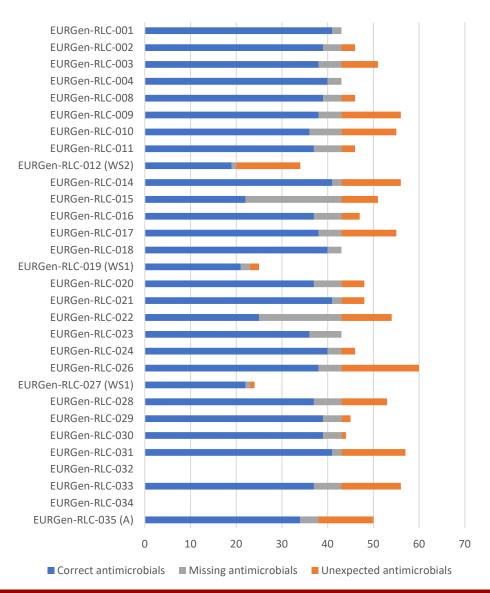
Material	Associated prediction of AMR profiles
EURGen-2023-01 ( <i>A. baumannii</i> )	<b>Expected</b> : Amikacin, ciprofloxacin, cefepime, ceftazidime, ceftazidime-avibactam, gentamicin, imipenem, meropenem, piperacillin-tazobactam, tobramycin
	Intrinsic: Aztreonam, fosfomycin
EURGen-2023-02 ( <i>E. coli</i> )	<b>Expected:</b> Amikacin, amoxicillin-clavulanic acid, ampicillin, aztreonam, cefepime, cefotaxime, ceftazidime, ceftazidime-avibactam, ciprofloxacin, ertapenem, <u>fosfomycin</u> , gentamicin, imipenem, meropenem, piperacillintazobactam, sulfamethoxazole, tobramycin, trimethoprim
	Expected non-mandatory: Colistin <sup>a</sup>
	Expected: Aztreonam, cefepime, cefotaxime, ceftazidime, ciprofloxacin
EURGen-2023-03 (K. pneumoniae)	Expected non-mandatory: Colistin <sup>b</sup>
(K. pheumomae)	Intrinsic: Ampicillin
EURGen-2023-04 ( <i>P. aeruginosa</i> )	<b>Expected:</b> Amikacin, cefepime, ceftazidime, ceftazidime-avibactam, ciprofloxacin, gentamicin, imipenem, meropenem, piperacillin-tazobactam, tobramycin

21-11-2023 Results from second EQA exercise Funded by the European Union









#### **Total submissions: 105**

- 10.5% submissions were fully correct (n=11)
- 63.8% had missing antimicrobials (n=67)
- 52.2% had unexpected antimicrobials (n=58)

29.5% Simultaneous (n=31)

Average score: 87.2%

None of participants achieved 100% of their max. possible score

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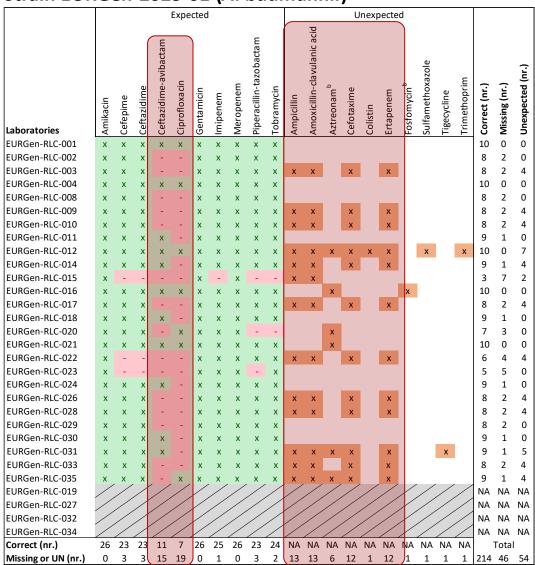
Strain	Total expected	Correct	Missing	Unexpected	
EURGen-2023-01	260	214 (82.3%)	46 (17.7%)	54 (个 20.8%)	←
EURGen-2023-02	486	430 (88.5%)	56 (11.5%)	2 (↑ 0.4%)	
EURGen-2023-03	130	120 (92.3%)	10 (7.7%)	43 (个 33.1%)	<b>←</b>
EURGen-2023-04	260	228 (87.7%)	32 (12.3%)	90 (个 34.6%)	<b>←</b>



# IN SILICO PREDICTION OF AMR PROFILES — SUBMITTED REULTS



#### Strain EURGen-2023-01 (A. baumannii)



- Missing from most submitted results
  - Ciprofloxacin (n=19)
    - Most participants used ResFinder which lacks database of AMR PMs for A. baumannii
  - Ceftazidime-avibactam (n=15)
    - Missing from bla<sub>NDM-1</sub> resistance profile in ResFinder database (but present for other genes)
- Reporting antimicrobials not included in the EQA
  - Ampicillin (n=13), Amoxicillin-clavulanic acid (n=13),
    Cefotaxime (n=12), Etrapenem (n=12)

#### Results must be evaluated critically:

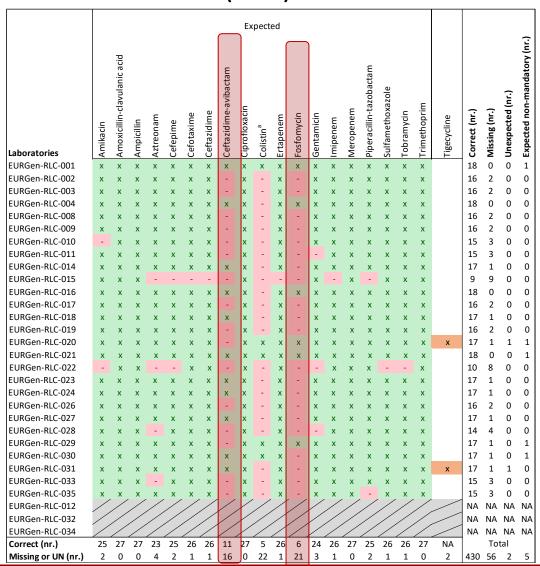
- Results can / should be confirmed with other tools
- Points to insufficient knowledge of genetic AMR mechanisms



# IN SILICO PREDICTION OF AMR PROFILES — SUBMITTED REULTS



#### Strain EURGen-2023-02 (E. coli)



- Missing from most submitted results
  - Ceftazidime-avibactam (n=16)
    - Missing from *bla*<sub>NDM-5</sub> R-profile in ResFinder database (but present for other genes)
  - Fosfomycin (n=21)
    - *glpT* E448K missing in ResFinder

#### Results must be evaluated critically:

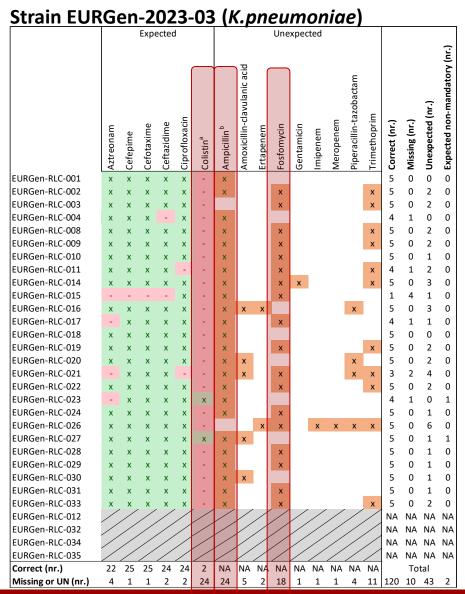
Results can / should be confirmed with other tools

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- Missing from most submitted results
  - Colistin (n=24)
    - Due to missing *mgrB*::IS1 mutation
- Reporting of Intrinsic resistance
  - Ampicillin (n=24)
  - Fosfomycin (n=18)

#### Results must be evaluated critically:

- Results can / should be confirmed with other tools
- Points to insufficient knowledge of genetic mechanisms of AMR

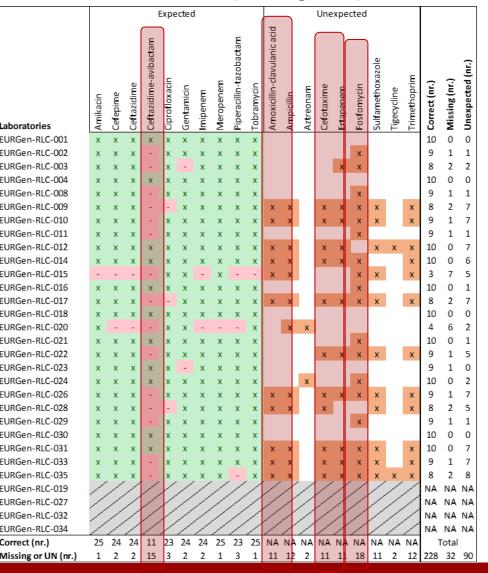




# IN SILICO PREDICTION OF AMR PROFILES – SUBMITTED REULTS



#### Strain EURGen-2023-04 (P. aeruginosa)



- Missing from most submitted results
  - Ceftazidime-avibactam (n=15)
    - Missing from *bla*<sub>NDM-5</sub> R-profile in ResFinder database (but present for other genes)
- Reporting antimicrobials not included in the EQA
  - Ampicillin (n=12), Amoxicillin-clavulanic acid (n=11), Cefotaxime (n=11), Etrapenem (n=11)
- Reporting incorrect resistance
  - Fosfomycin (n=18)
  - Not intrinsic in P. aruginosa (although carries fosA)

#### **Results must be evaluated critically:**

- Results can / should be confirmed with other tools
- Points to insufficient knowledge of genetic mechanisms of **AMR**



## **GENERAL RECOMMENDATIONS**



- For discrepancies due to misinterpretation of the EQA protocol and/or insufficient knowledge about certain genetic elements:
  - Laboratories should ensure sufficient knowledge about the genetic mechanisms mediating AMR and other important genetic elements;
  - Laboratories should ensure the use of multiple bioinformatics tools and databases for the detection of genetic determinants since bioinformatics tools and databases can be limited to the analysis of only a few bacterial species which contributes to the false-negative results;
  - Laboratories should be familiar with the bioinformatics tools they use, and the contents of the respective databases;
  - Laboratories should analyse their results critically and, when needed, perform confirmatory testing, to ensure that the information being reported is accurate.



# **GENERAL RECOMMENDATIONS**



- For discrepancies due to variations between the type of data and the chosen bioinformatics tools and databases:
  - Laboratories should implement verification steps such as using multiple bioinformatics approaches to confirm the obtained results;
  - Laboratories should communicate their suggestions, strange observations and potential problems to the curators of bioinformatics tools and databases;
  - Laboratories should be aware of differences between short-and long-read sequencing data and select the most adequate approach depending on their aims.
- For curators of bioinformatics tools for AMR surveillance:
  - Curators of widely-used bioinformatics tools and databases should try to improve the databases and regularly update the databases
  - Curators of bioinformatics tools and databases should engage in ongoing, active dialogue to ensure conformity between approaches;





# QUALITY CONTROL ANALYSIS OF SUBMITTED SEQUENCES

- METRICS INCLUDED IN QUALITY CONTROL ANALYSIS
- TOTAL ACHIEVED ILLUMINA SCORES
- RESULTS FROM QUALITY METRICS- ILLUMINA SEQUENCES
- NOTE ON OXFORD NANOPORE SEQUENCING





# **QC METRICS**

- Applied thresholds are metric specific
- Submitted genomes which deviate more than 10% from expected genomic size or does not have at least 95% of cgMLST alleles identified are expected to fail outright
- Coverage, Q-scores and proportion of mapping reads - are to some extent platform dependant
- Correlation groups 2 and 3 are noncorrelating groups, submission failing across both are indicative of large QC issue

Table 2. Overview of scoring criteria and their respective cut-off values and weight for new scheme.

Criteria	Correlation group	Minimal cut-off	Preferred cut-off	Score (%)	Score preferred (%)
cgMLST	1	95% match		15	
MLST	1	Must match		10	
Average coverage	1	>20x	>30x	2.5	5
Average Q-score R1	1	>25	>30	5	7.5
Average Q-score R2	1	>25	>30	5	7.5
Proportion of reads mapped to reference DNA (%)	1	>80%	>90%	2.5	5
Size of assembled genome	2	± 10 % of the reference genome size	Less than 3SD from mean of cleaned data set	5	7.5
Number of contigs > 200bp	2	Less than 3SD above median of cleaned data set		7.5	
Genomic coverage of minimal depth 10x	3	Less than 3SD below median of cleaned data set		7.5	
N50	3	Less than 3SD below mean of		7.5	

Table 2 from the Readme file for Illumina sequencing







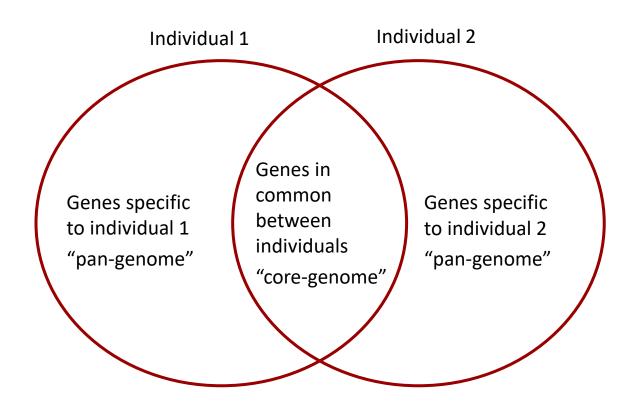
## **METRICS SHORT REFRESHER - MLST**

#### MLST

- Scheme of 7 specific core genes in isolates
- Each gene has a specific DNA sequence
- Any genetic variation is classified as a new allele
- The combination of these 7 unique alleles defines a sequence type
- Perfect match expected

#### cgMLST

- The concept above is applied to a predefined set of core genes expected to be present in every individual member of the species
- 95% match expected



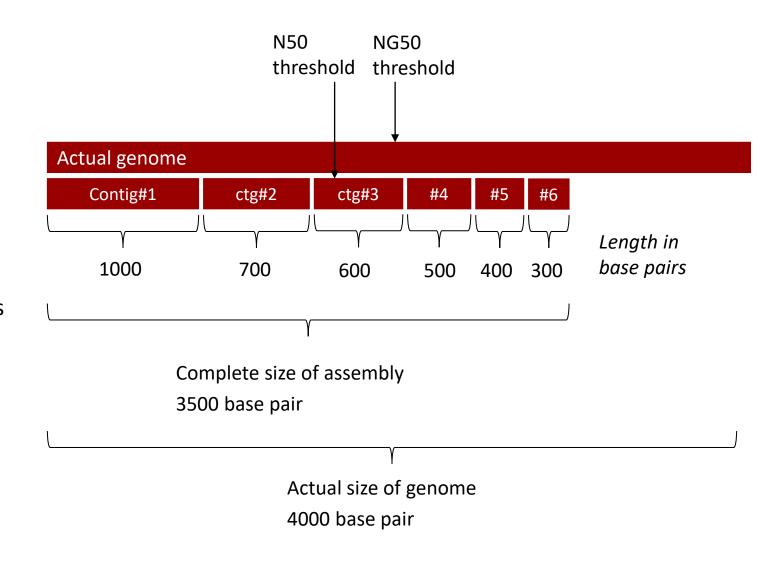
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#### **ASSEMBLY METRICS**



- Commonly used metrics
  - Number of contigs more than 200 bp
    - Here 6
  - N50
    - Adding lengths of contigs, going from longest to shortest, what is the minimal length included to reach 50% of the complete assembly size
  - Size of assembly compared to reference genome
    - Indicative of contamination or sequence quality
    - Here 87.5%
- Adjusted quality thresholds used for evaluation

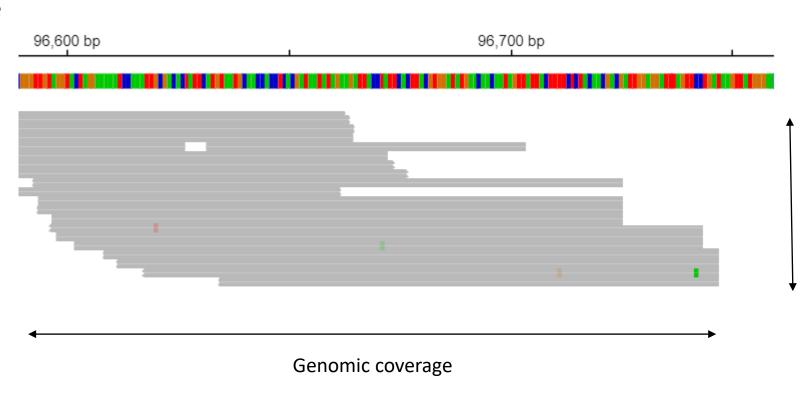


## **COVERAGE METRICS**

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Coverage/Depth

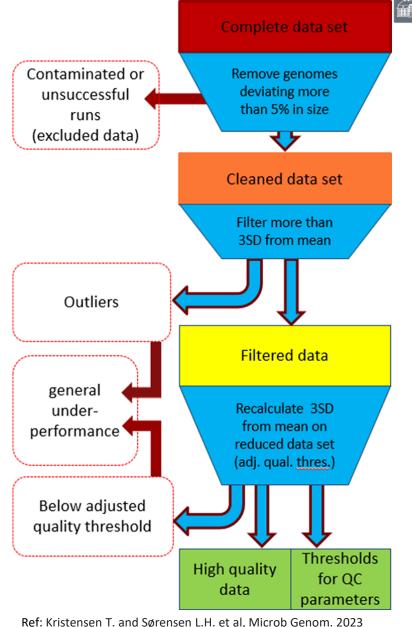
- Average coverage
  - On average, how many times is the genome covered by reads
  - Completely dependent on:
    - Sequencing yield
    - Size of the genome
  - Minimum of 20x, 30x prefered
- Genomic coverage of minimal depth
  10x
  - The breadth of the coverage of the reference genome
  - Low depth leads to greater uncertainty of the base call
  - Assembly is complicated by low coverage regions
  - Adjusted quality threshold





## **DEFINITION OF QC THRESHOLDS**

- Data was evaluated in three steps:
  - Initial exclusion
    - 10% > deviance from reference size
    - <95% cgMLST alleles
  - Outlier detection
    - More than 3 standard deviations from the mode
    - Mode is estimated either by mean or by median of the submitted genome
  - Setting adjusted quality threshold
    - Detected outliers were removed and standard deviations and mode was recalculated
    - This is done to remove data points which deviate by a large margin from the main body of data, as such outliers have a very large impact on the estimation of standard deviation and mode of the distribution



Aug;9(8):mgen001076. doi: 10.1099/mgen.0.001076. PMID: 37526643

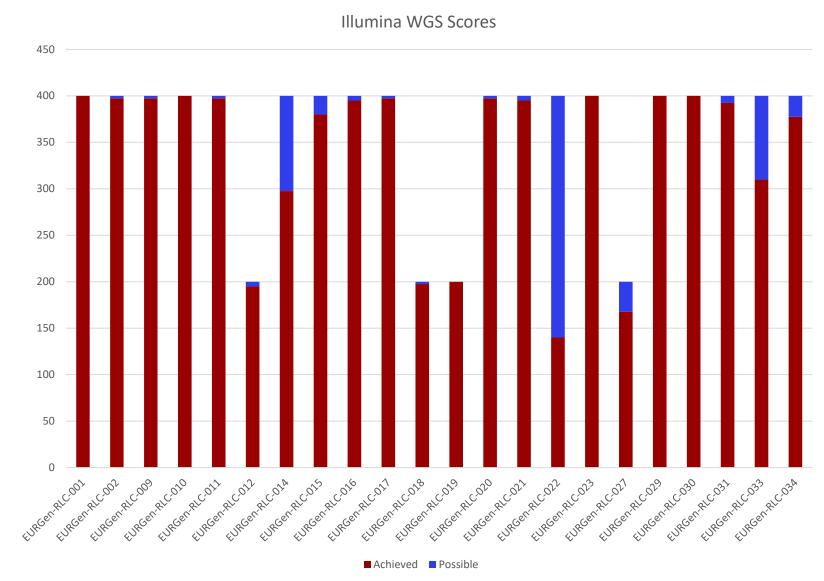






#### TOTAL ACHIEVED ILLUMINA SCORES

- We received Illumina WGS data from 22 laboratories
- Four participants submitted WGs for only two species, remaining submitted for all four available
- Three were less successful, mainly due to high variance in genome size compared to reference and lack of coverage

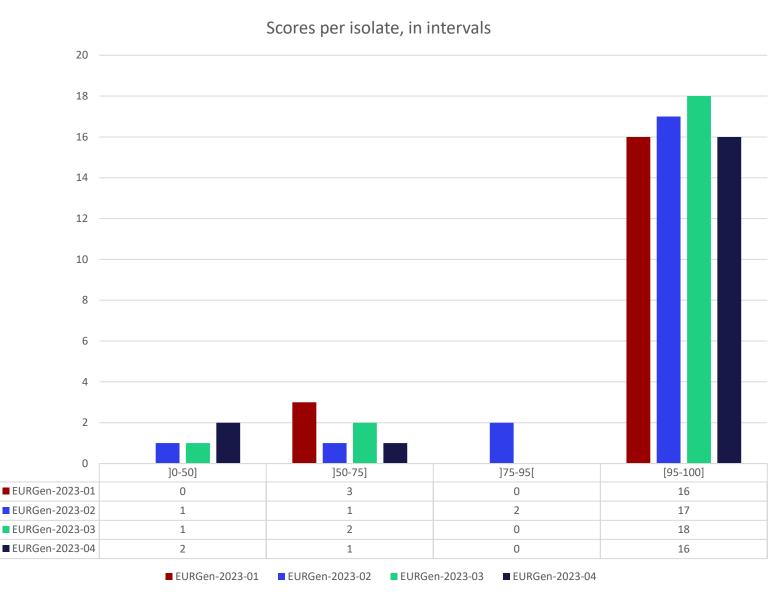




## TOTAL ACHIEVED ILLUMINA SCORES - SUMMARY



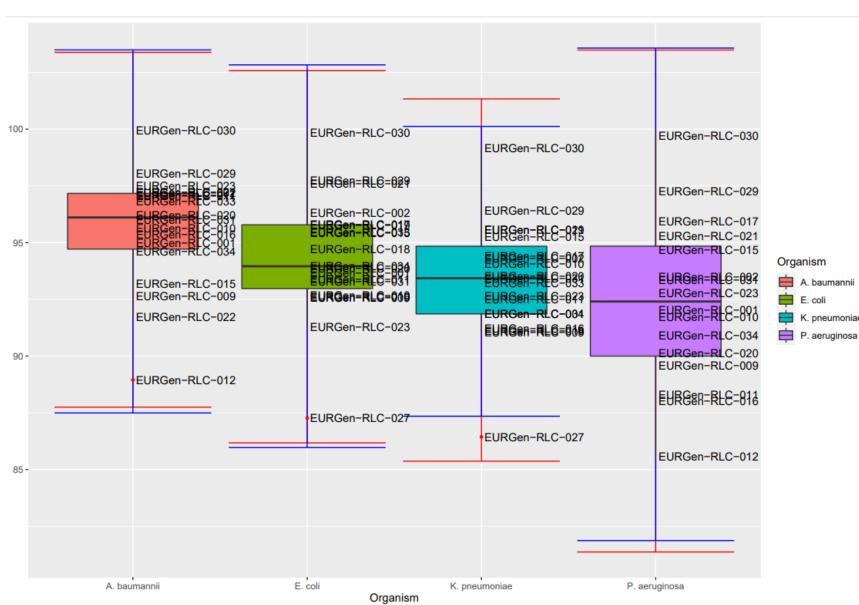
- For each species we received:
  - 19 A. baumannii
  - 21 *E. coli*
  - 21 K. pneumonia
  - 19 P. aeruginosa
- Issues were evenly found in all species, though A. baumannii and P. aeruginosa showed widest distributions in QC parameters





#### PROPORTION OF READS MAPPING TO REFERENCE

- Disregarding genomes removed in initial exclusion, all submission were above accepted threshold of 80%
- Most are above preferred threshold of 90%
- This metric is to some extent dependent on the sequencing platform



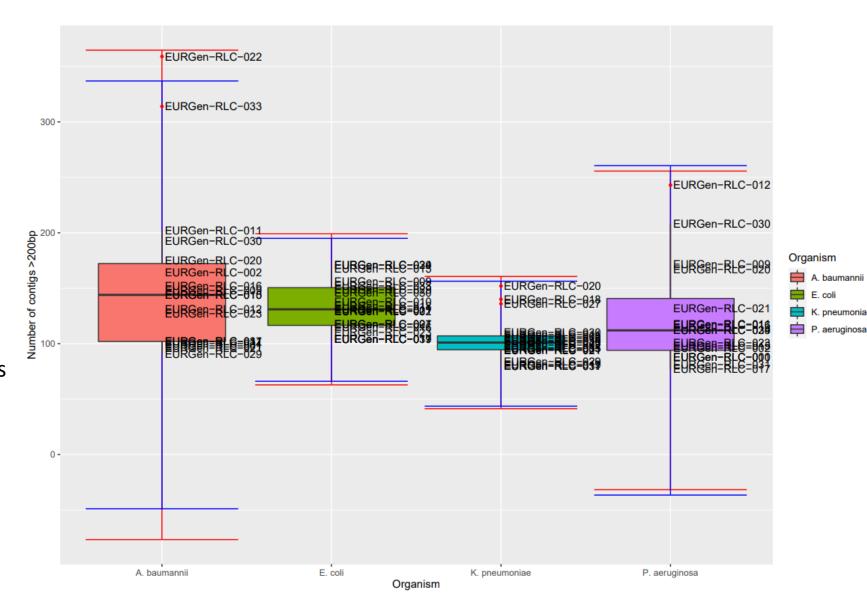
K. pneumoniae



# NUMBER OF CONTIGS > 200 bp



- Close distribution with some variance in *P. aeruginosa*
- Red whiskers are outlier threshold
- Blue whiskers are adj. qual. thresholds
- A couple of genomes with high counts in A. baumannii one of which is outside the adjusted quality threshold

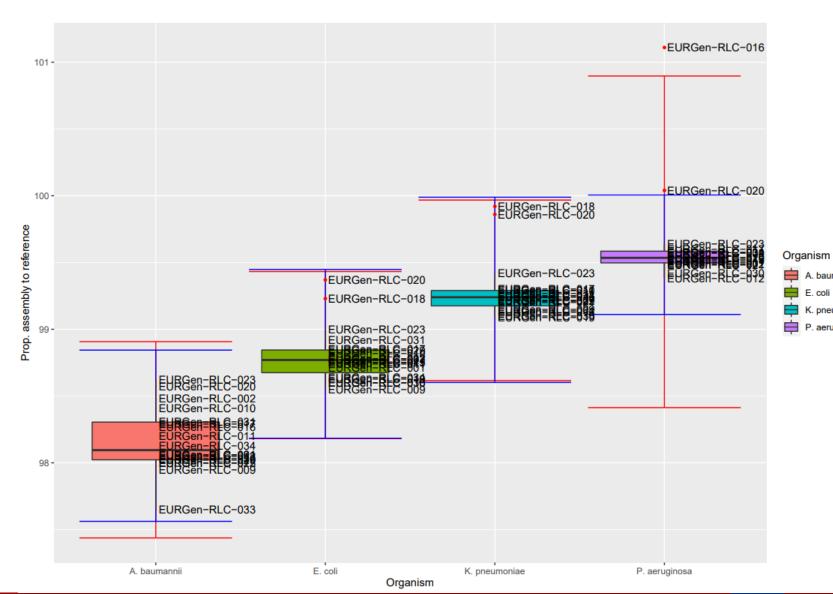




#### SIZE OF ASSEMBLY COMPARED TO REFERENCE



- Most assembly sizes are with expected thresholds
- Two submission for P. aeruginosa above general size of submissions, but notably only deviates 0.5-1.5%
- This metric is generally expected to be below 100% as this is not mapping, but the relative assembly size

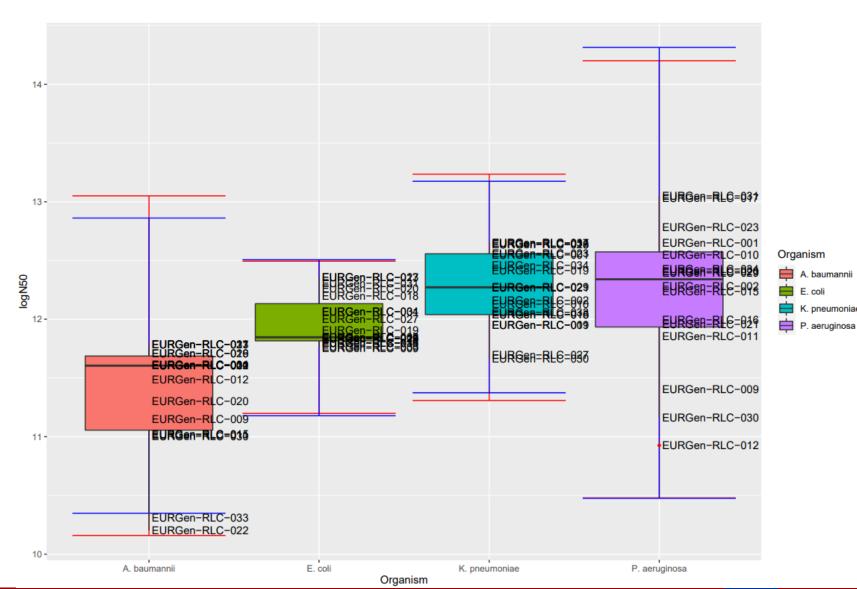


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#### **N50**

- Natural logarithmic transformation applied
- N50 generally shows comparable quality among participants
- In A. baumannii, two submission found outside adj. qual. threshold
- P. aeruginosa show a wide distribution comparable to other isolates



E. coli

K. pneumoniae



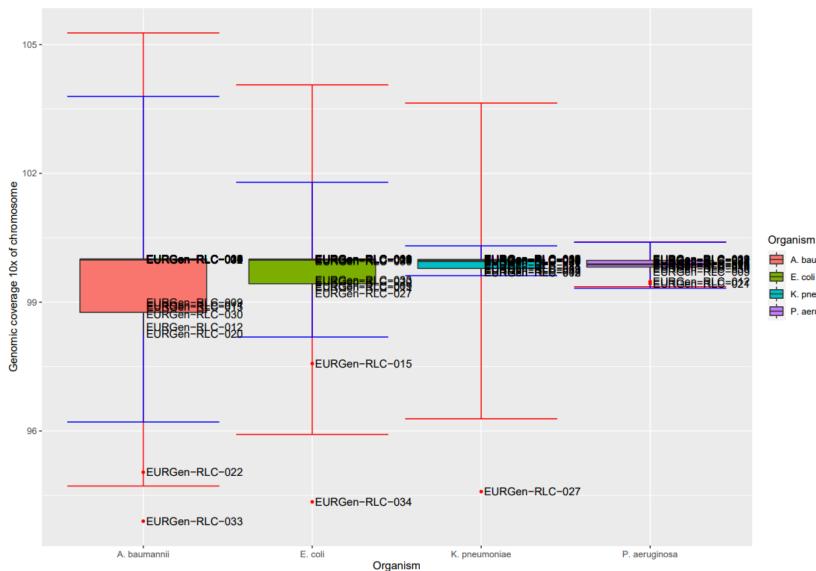
#### **GENOMIC COVERAGE OF MINIMAL 10X DEPTH**



 Most submitted genomes are above 99%, with notable exception of A. baumannii

 Three submission are outliers, two are below adj. qual. thresholds

• Having a high coverage of 10x is likely mainly influenced by the yield of the sequencing, but could also be due to bias in the workflow





## NOTE ON OXFORD NANOPORE SEQUENCING



- Filtered on length and quality before assembly
- Evaluation more individualized as quality control of ONT is less well understood
- Issues identified:
  - Achieving high enough yield for recommend coverage of 30x
  - Coverage of plasmids

Mapped to chromosome/plasmid_(N)	Number of reads mapping to the specific genomic component.
Total assembly size	Total number of base pairs in the assembly.
Number of contigs	The number of produced contigs compared to the number expected in the reference (chromosome + number of plasmids), shown as a fraction.
Number circularized	Number of contigs reported to be circularized by the assembler.
MLST	Identified MLST
Coverage of the reference genome/chromosome/plasmid_(N) (%)	Proportion of the reference genome, chromosome or plasmid (N) covered by reads (this cannot exceed 100%)
Coverage 20/30/40/50x of the reference genome/chromosome/plasmid_(N) (%)	Proportion of the reference genome, chromosome or plasmid N, covered by at least X times of reads. (This cannot exceed 100%).

Subset of Table 1 from the ONT readme file



21-11-2023

#### **FEEDBACK SURVEY**

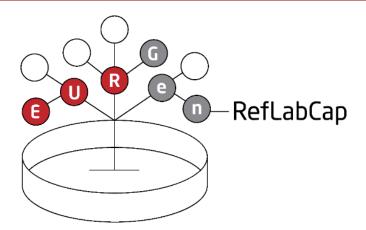


- Questions
  - How useful was this EQA to your laboratory? (scale:1-10)
  - Was the preliminary individual EQA evaluation report you received clear and useful? (Yes/No)
  - Did you take any corrective action(s)? (Yes/No)
  - Comment section for suggestions
- So far only 3 respondents
- The survey is still open (deadline= Nov 30)
- Via this link (<a href="https://ec.europa.eu/eusurvey/runner/EURGen-RefLabCap">https://ec.europa.eu/eusurvey/runner/EURGen-RefLabCap</a> EQA 2023 feedback survey October 2023 3e60685e-87a1-35eb-3bc9-486d41fd38a6)

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# Thank you on behalf of the EURGen-RefLabCap team

EURGen-RefLabCap@food.dtu.dk

