



SCIENTIFIC WEBINAR #1

GETTING STARTED WITH ONLINE SEQUENCE ANALYSIS BY CGE TOOLS

September 14th 2022
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- ✧ The concept of CGE tools
- ✧ Available tools
- ✧ Getting started
- ✧ Sequence types

Center for Genomic Epidemiology

- ❖ Collection of genomic tools with wide range of applications
- ❖ Some tools developed by CGE group, others are only linked
 - to have a simple and similar user interface
- ❖ Easy to use
 - Submit, get result by email, interpret
- ❖ Limited to one sequence/isolate at a time
- ❖ WGS – one sequence – numerous applications!
 - Does not (always) require coding skills & super-computers

Center for Genomic Epidemiology

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Overview of Services

Phenotyping

[ResFinder](#)

Identification of acquired antibiotic resistance genes.

[ResFinderFG](#)

Identification of functional metagenomic antibiotic resistance
determinants.

[LRE-finder](#)

Identif

[KmerFinder](#)

Identification of acquired antibiotic resistance genes using Kmers.

[PathogenFinder](#)

Prediction of a bacteria's pathogenicity towards human hosts.

Phylogeny

[MINTyper](#)

Identification of SNPs with automatic filtering, masking and site
validation together with inferred phylogeny based on both long and
short sequencing data.

NDtree constructs phylogenetic trees from Single-End or Pair-End
FASTQ files.

[Evergreen](#)

The aim of the Center for Genomic Epidemiology (CGE) is to provide access to bioinformatics resources also for those with limited experience and thereby allow all countries, institutions and individuals to take advantage of the sequencing technologies.

- ❖ **Phenotyping**
- ❖ **Typing**
- ❖ **Phylogeny**
- ❖ **Metagenomics**
- ❖ **PCR tool**

Sequence contamination
Species determination
Subspecies typing
Resistance genes and mutations
Outbreak/cluster analysis



- ❖ **Others:**
 - User-defined data bases
 - Identify bacteriophage sequences in metagenomic data
 - Identify the bacterial host of phage genome
 - ...
 - ...

❖ Phenotyping

- Antimicrobial resistance
 - **ResFinder**
 - ResFinderFG
 - identifies a resistance phenotype based on a functional metagenomic antibiotic resistance determinants database
 - **Kmer resistance**
 - LRE-Finder
 - Identification of genes and mutations leading to linezolid resistance



❖ Species identification

- **KmerFinder** (kmer-based)
- SpeciesFinder (16S ribosomal DNA)

❖ Typing

- MLST
- cgMLSTFinder
 - *Campylobacter, Clostridium, E. coli, Listeria, Salmonella, Yersinia*
- pMLST
- Serotyping (*E. coli, P. aeruginosa, Salmonella*)
- PlasmidFinder
- MGE (Mobile genetic elements)

EXAMPLE: KMER-FINDER (SPECIES ID)

KmerFinder 3.2

Service **Instructions** Output Article a

Software version: 3.0.2 (2020-10-30)

Database version: (2022-04-01)

The database can be downloaded [here](#)


Select database

Archaea

Upload file(s)

To input the sequences, upload a single FASTA file, or one/two FASTQ file(s), or one interleaved FASTQ file on your local disk by using the applet below. Both assembled genome (in FASTA format) and raw reads single end or paired end (in FASTQ format) are supported. Gzipped FASTA/FASTQ files are also supported.

If you get an "Access forbidden. Error 403": Make sure the start of the web address is https and not just http. Fix it by clicking [here](#).

 Choose File(s)

Name

Size

Progress

Status

 Upload

 Remove

upload a single FASTA file, or one/two FASTQ file(s),

Select database

Archaea

Archaea

Bacteria organisms

Fungi

Protozoa

Bacteria type strains

Viral

Center for Genomic Epidemiology

Your job is being processed

Wait here to watch the progress of your job, or fill in the form below to get an email message upon completion.

To get notified by email:

This page will update itself automatically.

Link to result output can be shared
– valid for a few days or a week

KmerFinder-3.2 Server - Results

KmerFinder 3.2 results:

Template	Num	Score	Expected	Template_length	Query_Coverage	Template_Coverage
NZ_LR134222.1 Escherichia coli strain NCTC11129 genome assembly, chromosome: 1	134	143410	5	165253	89.50	87.68
NZ_CP069706.1 Escherichia coli strain ECY44 chromosome, complete genome	20183	5165	47	157604	3.22	3.26
NZ_CP034787.1 Escherichia coli strain ECCNB20- 2 chromosome, complete genome	1660	2551	48	158345	1.59	1.64

- ✦ Good coverage of Query/Template = good match to data base isolate
- ✦ A result with several hits is usual
- ✦ Check for contamination from other species

❖ Explanations on how to interpret output on website

KmerFinder 3.2

[Service](#)[Instructions](#)[Output](#)[Article abstract](#)[Citations](#)

Graphical output example and explanation

Once the KmerFinder server has finished running the job you submitted, it will display an output similar to the example below.

Explanation of the standard output

An example output of the KmerFinder using the **standard** scoring method is given in the image below. This example was generated using an assembled genome of a *Citrobacter freundii* strain.

KmerFinder 3.0 results:

Template	Num	Score	Expected	Template length
NZ_CP016952.1 Citrobacter freundii strain SL151 chromosome, complete genome	1723	127691	21	168352

Explanation of the columns in standard and extended output

The following contains a brief explanation of all columns of the output including th

Template: shows the accession numbers or name of the template sequences

Assembly: RefSeq assembly accession ID

Num: is the sequence number of accession entry in the KmerFinder database

Score: is the total number of matching Kmers between the query and the template

Expected: is the expected score, i.e. the expected total number of matching Kmers

Template length: is the number of Kmers in the template

Query_Coverage [%]: is the percentage of input query Kmers that match the tem

Template_Coverage [%]: is the template coverage.

Depth: is the number of matched kmers in the query sequence divided by the tota
sequencing depth.

ResFinder 4.1

Service Instructions Output Article abstract Citations Over

ResFinder identifies acquired genes and/or finds chromosomal mutations mediated in total or partial DNA sequence of bacteria.

ResFinder and PointFinder software: [\(2022-08-08\)](#)

ResFinder database: [EFSA_2021 \(2022-05-24\)](#)

PointFinder database: [\(2021-02-01\)](#)

For analysis part of EFSA, go to [ResFinder-EFSA](#)

Chromosomal point mutations ☐

Acquired antimicrobial resistance genes ☐

Select species

Campylobacter spp.*

*Chromosomal point mutation database exists

Select type of your reads

Assembled Genome/Contigs

Chromosomal point mutations ☒

Select threshold for %ID

90 %

Select minimum length

60 %

☐ Show unknown mutations, not found in the database

Acquired antimicrobial resistance genes ☒

Select Antimicrobial configuration

Select multiple items, with Ctrl-Click (or Cmd-Click on Mac) - as default all databases are selected

Aminoglycoside
Beta-lactam
Colistin
Disinfectant
Fluoroquinolone
Fosfomycin

Select threshold for %ID

90 %

Select minimum length

60 %

ResFinder 4.1

Service

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Chromosomal point mutations ☐

Acquired antimicrobial resistance genes ☐

Select species

Campylobacter spp.*

*Chromosomal point mutation database exists

Select type of your reads

Assembled Genome/Contigs

Select species

Campylobacter spp.*

Campylobacter spp.*

Campylobacter jejuni*

Campylobacter coli*

Escherichia coli*

Salmonella spp.*

Plasmodium falciparum*

Neisseria gonorrhoeae*

Mycobacterium tuberculosis*

Enterococcus faecalis*

Enterococcus faecium*

Klebsiella*

Helicobacter pylori*

Staphylococcus aureus*

Other

Select type of your reads

Assembled Genome/Contigs

Assembled Genome/Contigs

454 - single end reads

454 - paired end reads

Illumina - single end reads

Illumina - paired end reads

Ion Torrent

SOLiD - single end reads

SOLiD - paired end reads

SOLiD - mate pair reads

- ❖ Species specific overview of AMR genes present
 - Prediction of class AND phenotype

escherichia coli		complete		
Antimicrobial	Class	WGS-predicted phenotype	Genetic background	
amikacin	aminoglycoside	No resistance		
tigecycline	tetracycline	No resistance		
tobramycin	aminoglycoside	No resistance		
cefepime	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)	
chloramphenicol	phenicol	Resistant	floR (floR_AF118107)	
piperacillin+tazobactam	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)	
cefoxitin	beta-lactam	No resistance		
ampicillin	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550), blaTEM-1B (blaTEM-1B_AY458016)	
ampicillin+clavulanic acid	beta-lactam	No resistance		
cefotaxime	beta-lactam	No resistance		
ciprofloxacin	fluoroquinolone	Resistant	gyrA (p.S83L)	
colistin	polymyxin	Resistant	mcr-1.1 (mcr-1.1_KP347127)	
sulfamethoxazole	folate pathway antagonist	Resistant	sul2 (sul2_HQ840942)	
imipenem	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)	

- Gene variants and non-perfect matches
- Multiple matches in database – multiple matches in output

escherichia coli complete			
Antimicrobial	Class	WGS-predicted phenotype	Genetic background
amikacin	aminoglycoside	No resistance	
tobramycin	aminoglycoside	No resistance	
gentamicin	aminoglycoside	No resistance	
cefepime	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550), blaTEM-29 (blaTEM-29_DQ269440)
piperacillin+tazobactam	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550), blaTEM-122 (blaTEM-122_AY307100)
cefoxitin	beta-lactam	No resistance	
ampicillin	beta-lactam	Resistant	blaTEM-1B (blaTEM-1B_AY458016), blaOXA-162 (blaOXA-162_GU197550), blaTEM-29 (blaTEM-29_DQ269440), blaTEM-122 (blaTEM-122_AY307100), blaTEM-55 (blaTEM-55_DQ286729), blaTEM-141 (blaTEM-141_AY956335), blaTEM-57 (blaTEM-57_FJ405211), blaTEM-1C (blaTEM-1C_FJ560503), blaTEM-135 (blaTEM-135_GQ896333)
ampicillin+clavulanic acid	beta-lactam	Resistant	blaTEM-122 (blaTEM-122_AY307100)
cefotaxime	beta-lactam	Resistant	blaTEM-29 (blaTEM-29_DQ269440)
imipenem	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)
ertapenem	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)
ceftazidime	beta-lactam	Resistant	blaTEM-29 (blaTEM-29_DQ269440)
temocillin	beta-lactam	No resistance	
meropenem	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)
ciprofloxacin	fluoroquinolone	Resistant	gyrA (p.S83L)
nalidixic acid	fluoroquinolone	Resistant	gyrA (p.S83L), gyrA (p.D87N)

- Beta-lactam resistance - ampicillin

- TEM-1B, TEM-1C, blaTEM-29, blaTEM-122, blaTEM-55, blaTEM-57, blaTEM-135, blaTEM-141

EXTENDED OUTPUT

blaTEM-122	99.8838559814	861/861	1..861	NODE_96_lengt h_8473_cov_26. 387970	7452..8312	amoxicillin, amoxi cillin+clavulanic acid, ampicillin, a mpicillin+clavulan ic acid, piperacillin, p iperacillin+tazoba ctam, ticarcillin, tic arcillin+clavulanic acid
blaTEM-55	99.8838559814	861/861	1..861	NODE_96_lengt h_8473_cov_26. 387970	7452..8312	amoxicillin, ampici llin, cephalothin, pi peracillin, ticarcilli n
blaTEM-209	99.8838559814	861/861	1..861	NODE_96_lengt h_8473_cov_26. 387970	7452..8312	unknown beta- lactam
blaTEM-1B	99.8838559814	861/861	1..861	NODE_96_lengt h_8473_cov_26. 387970	7452..8312	amoxicillin, ampici llin, cephalothin, pi peracillin, ticarcilli n
blaTEM-141	99.8838559814	861/861	1..861	NODE_96_lengt h_8473_cov_26. 387970	7452..8312	amoxicillin, ampici llin, cephalothin, pi peracillin, ticarcilli n
blaOXA-162	100.0	798/798	1..798	NODE_163_leng th_2231_cov_6.1 64122	26..823	amoxicillin, ampici llin, cefepime, erta penem, imipenem meropenem, piper racillin, piperacilli n+tazobactam
blaTEM-57	99.8838559814	861/861	1..861	NODE_96_lengt h_8473_cov_26. 387970	7452..8312	amoxicillin, ampici llin, cephalothin, pi peracillin, ticarcilli n
blaTEM-29	99.8838559814	861/861	1..861	NODE_96_lengt h_8473_cov_26. 387970	7452..8312	amoxicillin, ampici llin, aztreonam, ce fepime, cefotaxim e, ceftazidime, ceft riaxone, piperacilli n, ticarcillin

All the TEM- genes
are in fact the same
gene

– placed on
NODE_96 in
position 7452-8312.

None of them have
a perfect match, so
it is likely a new
variant, bad
sequence or at least
something that
wasn't recognized
by the database.

Try running on raw
reads (fastq) instead
of contigs (fasta)

- Almost all the CGE tools are available for Illumina reads/fastq files

Tool	Result
KmerFinder	Species identification/confirmation and check for contamination
ResFinder (Incl. PointFinder)	Identification of acquired resistance genes and point mutations, phenotype prediction
PlasmidFinder/MGEFinder	Predicting presence of plasmids/relation to resistance genes
MLST	Sub-species level typing and check for contamination
CSIPhylogeny	Cluster detection and SNP calling
...etc....	

- ❖ Some tools can also analyse ONT MinION data, including:

Tool	Result
KmerFinder	Species identification/confirmation and check for contamination
KmerResistance	Identification of acquired resistance genes
MLST	Sub-species level typing and check for contamination
MinTyper	Cluster detection and SNP calling

- ❖ Subtyping and cluster analysis will be presented on the virtual multidisciplinary exercise on Sept 26th and Oct 10th
 - More information about this next week

- ✧ For local installation
 - Download tools and databases at

<https://bitbucket.org/genomicepidemiology/>

https://bitbucket.org/genomicepidemiology/resfinder_db/

<https://bitbucket.org/genomicepidemiology/kmerfinder/src/master/>

Installation

Download the ResFinder database by cloning the database:

```
git clone https://git@bitbucket.org/genomicepidemiology/resfinder_db.git
```

The database can be used with BLAST as-is.

Main site:

<https://www.genomicepidemiology.org/>

<http://www.genomicepidemiology.org/services/>

<https://cge.food.dtu.dk/services/MLST/>

Please note that the links
have changed recently

<https://cge.food.dtu.dk/services/KmerFinder/>

<https://cge.food.dtu.dk/services/ResFinder/>

<https://cge.food.dtu.dk/services/MINTyper/>

<https://cge.food.dtu.dk/services/CSIPhylogeny/>

Thank you for the attention!