







#### **SCIENTIFIC WEBINAR #1**

# GETTING STARTED WITH ONLINE SEQUENCE ANALYSIS BY CGE TOOLS

September 14<sup>th</sup> 2022 Jette S. Kjeldgaard



## **AGENDA**



- The concept of CGE tools
- Available tools
- Getting started
- Sequence types

## Center for Genomic Epidemiology



## THE CONCEPT OF CGE TOOLS?



- Collection of genomic tools with wide range of applications
- Some tools developed by CGE group, others are only linked
  - to have a simple and similar user interface
- : Easy to use
  - Submit, get result by email, interpret
- Limited to one sequence/isolate at a time
- ▶ WGS one sequence numerous applications!
  - Does not (always) require coding skills & super-computers





## Center for Genomic Epidemiology

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#### Overview of Services

### Phenotyping

#### ResFinder

Identification of acquired antibiotic resistance genes.

#### **ResFinderFG**

Identifcation of functional metagenomic antibiotic resistance

The aim of the Center for Genomic Epidemiology (CGE) is to

provide access to bioinformatics resources also for those with limited experience and thereby allow all countries, institutions and individuals to take advantage of the sequencing technologies.

Kmer

Identification of acquired antibiotic resistance genes using Kmers.

#### <u>PathogenFinder</u>

Prediction of a bacteria's pathogenicity towards human hosts.

#### **Phylogeny**

#### <u>MINTyper</u>

Identification of SNPs with automatic filtering, masking and site validation together with inferred phylogeny based on both long and short sequencing data.

NDtree constructs phylogenetic trees from Single-End or Pair-End FASTQ files.

#### **Evergreen**

n and

the high

## **CGE TOOLS COLLECTION - OVERVIEW**



- Phenotyping
- Typing
- Phylogeny
- Metagenomics
- **PCR** tool

Sequence contamination
Species determination
Subspecies typing
Resistance genes and
mutations
Outbreak/cluster analysis



#### • Others:

- User-defined data bases
- Identify bacteriophage sequences in metagenomic data
- Identify the bacterial host of phage genome
- ...
- ..

## CGE TOOLS PHENOTYPING - AMR



## Phenotyping

- Antimicrobial resistance
  - ResFinder
  - ResFinderFG
    - identifies a resistance phenotype based on a functional metagenomic antibiotic resistance determinants database
  - Kmer resistance
  - LRE-Finder
    - Identification of genes and mutations leading to linezolid resistance

## **CGE TOOLS TYPING**



### Species identification

- KmerFinder (kmer-based)
- SpeciesFinder (16S ribosomal DNA)

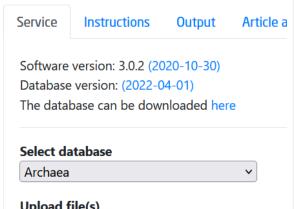
## Typing

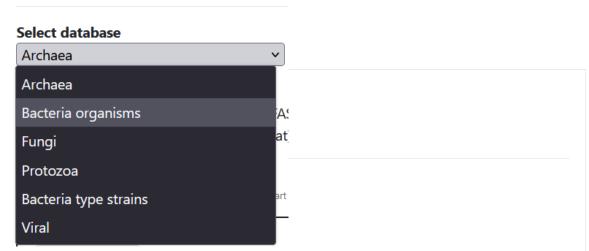
- MLST
- cgMLSTFinder
  - Campylobacter, Clostridum, E. coli, Listeria, Salmonella, Yersinia
- pMLST
- Serotyping (E. coli, P. aeruginosa, Salmonella)
- PlasmidFinder
- MGE (Mobile genetic elements)

## **EXAMPLE: KMER-FINDER (SPECIES ID)**









#### Upload file(s)

To input the sequences, upload a single FASTA file, or one/two FASTQ file(s), or one interleaved FASTQ file on your local disk by using the applet below. Both assembled genome (in FASTA format) and raw reads single end or paired end (in FASTQ format) are supported. Gzipped FASTA/FASTQ files are also supported.

If you get an "Access forbidden. Error 403": Make sure the start of the web adress is https and not just http. Fix it by clicking here.



## **EXAMPLE:** KMER-FINDER (SPECIES ID)



## Center for Genomic Epidemiology

## Your job is being processed

Wait here to watch the progress of your job, or fill in the form below to get an email message upon completion.			
To get notified by email:	Notify me via email		
This page will update itself automatically.			

Link to result output can be sharedvalid for a few days or a week

## SPECIES ID - MATCH TO DATABASE



#### KmerFinder-3.2 Server - Results

#### KmerFinder 3.2 results:

Template	Num	Score	Expected	Template_length	Query_Coverage	Template_Coverage
NZ_LR134222.1 Escherichia coli strain NCTC11129 genome assembly, chromosome: 1	134	143410	5	165253	89.50	87.68
NZ_CP069706.1 Escherichia coli strain ECY44 chromosome, complete genome	20183	5165	47	157604	3.22	3.26
NZ_CP034787.1 Escherichia coli strain ECCNB20- 2 chromosome, complete genome	1660	2551	48	158345	1.59	1.64

- Good coverage of Query/Template = good match to data base isolate
- A result with several hits is usual
- Check for contamination from other species

## INTERPRETATION OF KMER-FINDER RESULT



Explanations on how to interpret output on website

#### KmerFinder 3.2

Article abstract Citation	nstructions	Service
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#### Graphical output example and explanation

Once the KmerFinder server has finished running the job you submitted, it will display an output similar to the example below.

#### Explanation of the standard output

An example output of the KmerFinder using the **standard** scoring method is given in the image below. This example was generated using an assembled genome of a Citrobacter freundii strain.

#### KmerFinder 3.0 results:

Template	Num	Score	Expected	Template length
NZ_CP016952.1 Citrobacter freundii strain SL151 chromosome, complete genome	1723	127691	21	168352

#### Explanation of the columns in standard and extended output

The following contains a brief explanation of all columns of the output including the

**Template:** shows the accession numbers or name of the template sequences

Assembly: RefSeq assembly accession ID

Num: is the sequence number of accession entry in the KmerFinder database

Score: is the total number of matching Kmers between the query and the template

Expected: is the expected score, i.e.the expected total number of matching Kmers

**Template length:** is the number of Kmers in the template

Query\_Coverage [%]: is the percentage of input query Kmers that match the tem;

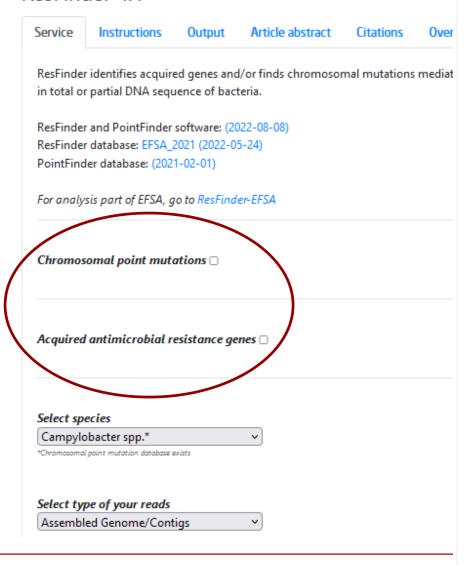
**Template\_Coverage** [%]: is the template coverage.

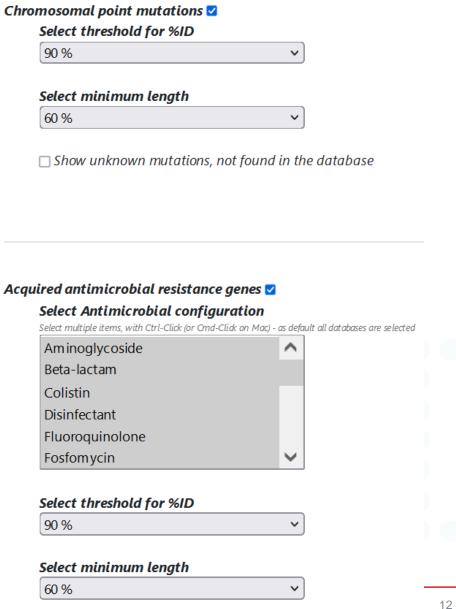
**Depth:** is the number of matched kmers in the query sequence divided by the total sequencing depth.

## **EXAMPLE: RESFINDER**



#### ResFinder 4.1

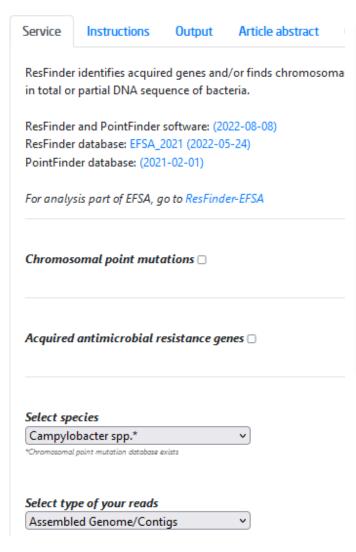




## **EXAMPLE: RESFINDER**

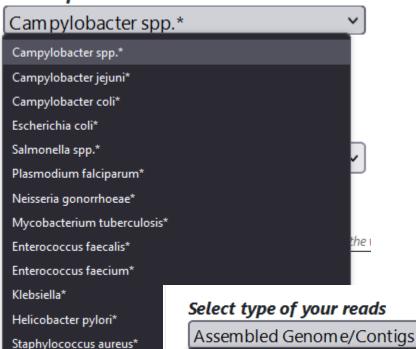


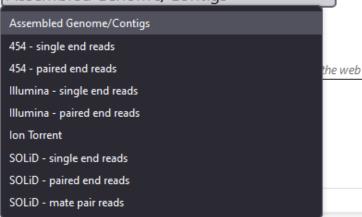
#### ResFinder 4.1



#### Select species

Other





## **RESFINDER OUTPUT**



- Species specific overview of AMR genes present
  - Prediction of class AND phenotype

escherichia coli comple	te		
Antimicrobial	Class	WGS-predicted phenotype	Genetic background
amikacin	aminoglycoside	No resistance	
tigecycline	tetracycline	No resistance	
tobramycin	aminoglycoside	No resistance	
cefepime	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)
chloramphenicol	phenicol	Resistant	floR (floR_AF118107)
piperacillin+tazobactam	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)
cefoxitin	beta-lactam	No resistance	
ampicillin	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550), blaTEM-1B (blaTEM-1B_AY458016)
ampicillin+clavulanic acid	beta-lactam	No resistance	
cefotaxime	beta-lactam	No resistance	
ciprofloxacin	fluoroquinolone	Resistant	gyrA (p.S83L)
colistin	polymyxin	Resistant	mcr-1.1 (mcr-1.1_KP347127)
sulfamethoxazole	folate pathway antagonist	Resistant	sul2 (sul2_HQ840942)
imipenem	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)

## **OUTPUT QUALITY DEPENDENT ON INPUT**



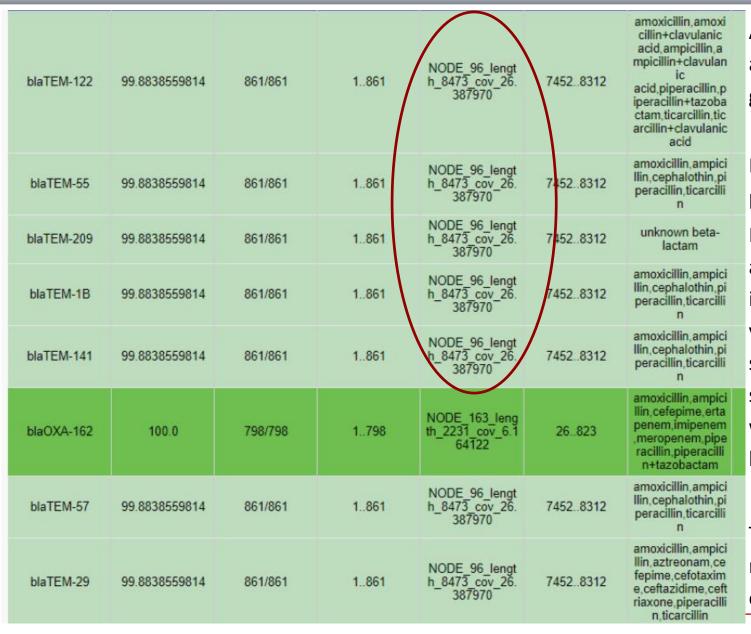
- Gene variants and non-perfect matches
- Multiple matches in database multiple matches in output

escherichia coli complete					
Antimicrobial	Class	WGS-predicted phenotype	Genetic background		
amikacin	aminoglycoside	No resistance			
tobramycin	aminoglycoside	No resistance			
gentamicin	aminoglycoside	No resistance			
cefepime	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550), blaTEM-29 (blaTEM-29_DQ269440)		
piperacillin+tazobactam	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550), blaTEM-122 (blaTEM-122_AY307100)		
cefoxitin	beta-lactam	No resistance			
ampicillin	beta-lactam	Resistant	blaTEM-1B (blaTEM-1B_AY458016), blaOXA-162 (blaOXA-162_GU197550), blaTEM-29 (blaTEM-29_DQ269440), blaTEM-122 (blaTEM-122_AY307100), blaTEM-55 (blaTEM-55_DQ286729), blaTEM-141 (blaTEM-141_AY956335), blaTEM-57 (blaTEM-57_FJ405211), blaTEM-1C_FJ560503), blaTEM-135 (blaTEM-135_GQ896333)		
ampicillin+clavulanic acid	beta-lactam	Resistant	blaTEM-122 (blaTEM-122_AY307100)		
cefotaxime	beta-lactam	Resistant	blaTEM-29 (blaTEM-29_DQ269440)		
imipenem	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)		
ertapenem	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)		
ceftazidime	beta-lactam	Resistant	blaTEM-29 (blaTEM-29 DQ269440)		
temocillin	beta-lactam	No resistance			
meropenem	beta-lactam	Resistant	blaOXA-162 (blaOXA-162_GU197550)		
ciprofloxacin	fluoroquinolone	Resistant	gyrA (p.S83L)		
nalidixic acid	fluoroquinolone	Resistant	gyrA (p.S83L), gyrA (p.D87N)		

- ▶ Beta-lactam resistance ampicillin
  - TEM-1B, TEM-1C, 1TEM-29, TEM-122, TEM-55, TEM-57, TEM-135, TEM-141

## **EXTENDED OUTPUT**





All the TEM- genes are in fact the same gene

placed on
NODE\_96 in
position 7452-8312.
None of them have
a perfect match, so
it is likely a new
variant, bad
sequence or at least
something that
wasn't recognized
by the database.

Try running on raw reads (fastq) instead of contigs (fasta)

## ILLUMINA FASTQ/FASTA



. Almost all the CGE tools are available for Illumina reads/fasta files

Tool	Result
KmerFinder	Species identification/confirmation and check for contamination
ResFinder (Incl. PointFinder)	Identification of acquired resistance genes and point mutations, phenotype prediction
PlasmidFinder/MGEFinder	Predicting presence of plasmids/relation to resistance genes
MLST	Sub-species level typing and check for contamination
CSIPhylogeny	Cluster detection and SNP calling
etc	

## **ONT MINION**



Some tools can also analyse ONT MinION data, including:

Tool	Result
KmerFinder	Species identification/confirmation and check for contamination
KmerResistance	Identification of acquired resistance genes
MLST	Sub-species level typing and check for contamination
MinTyper	Cluster detection and SNP calling

- Subtyping and cluster analysis will be presented on the virtual multidisciplinary exercise on Sept 26<sup>th</sup> and Oct 10<sup>th</sup>
  - More information about this next week

## **AVAILABILITY OF TOOLS**



- For local installation.
  - Download tools and databases at

## https://bitbucket.org/genomicepidemiology/

https://bitbucket.org/genomicepidemiology/resfinder\_db/

https://bitbucket.org/genomicepidemiology/kmerfinder/src/master/

#### Installation

Download the ResFinder database by cloning the database:

git clone https://git@bitbucket.org/genomicepidemiology/resfinder\_db.git

The database can be used with BLAST as-is.



## LINKS



Main site:

https://www.genomicepidemiology.org/

http://www.genomicepidemiology.org/services/

https://cge.food.dtu.dk/services/MLST/

Please note that the links have changed recently

https://cge.food.dtu.dk/services/KmerFinder/

https://cge.food.dtu.dk/services/ResFinder/

https://cge.food.dtu.dk/services/MINTyper/

https://cge.food.dtu.dk/services/CSIPhylogeny/

## Thank you for the attention!